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Genetic Testing Summary

Enclosed are the genetic testing results for

CB 556

No amount of genetic testing can guarantee that a child will not be affected with a genetic condition. Genetic testing can inform you of the likelihood of passing on the genetic conditions that are tested for, but it cannot eliminate the risk of passing on any genetic condition.

The genetic conditions Cryobio tests for are inherited in an autosomal recessive manner. This means that the child would have to inherit a genetic mutation from both the sperm source and the egg source to be affected with the condition. When both the sperm source and the egg source have undergone genetic carrier screening and the test results are negative, the risk of a child being affected with the conditions tested for is significantly reduced, but it cannot be completely eliminated.

All recipients should discuss both their own risk for passing on genetic conditions and whether they would benefit from genetic counseling and testing with their health care provider. Before using a donor that is a carrier for a specific recessive genetic condition or conditions, we strongly recommend that the recipient (or egg source, if different) consider genetic counseling and testing to determine if they are a carrier for the same genetic condition or conditions as the donor.

Screening and testing have changed dramatically over the years, and so the screening and testing done on each donor may vary depending on the testing that was in place when he was actively in Cryobio's donor program. Earlier donors may not have had as extensive testing as later donors. Screening and testing may change again in the future, so please review the results each time before ordering as both the testing done and the results may change.

Patient Information

Name: Cb 556
 Date of Birth: [REDACTED]
 Sema4 ID [REDACTED]
 Client ID [REDACTED]
 Indication: Carrier Testing

Specimen Information

Specimen Type: Blood
 Date Collected: [REDACTED]
 Date Received: [REDACTED]
 Final Report: [REDACTED]

Referring Provider

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Expanded Carrier Screen (283)

Number of genes tested: 283

SUMMARY OF RESULTS AND RECOMMENDATIONS

| ⊕ Positive | ⊖ Negative |
|--|---|
| <p>Carrier of Beta-Globin-Related Hemoglobinopathies (AR) Associated gene(s): <i>HBB</i> Variant(s) Detected: c.208G>A, p.G70S (Hb City of Hope), Likely Pathogenic, Heterozygous (one copy)</p> <p>Carrier of Congenital Adrenal Hyperplasia due to 21-Hydroxylase Deficiency (AR) Associated gene(s): <i>CYP21A2</i> Variant(s) Detected: c.841G>T, p.V281L, Pathogenic, Heterozygous (one copy)</p> <p>Carrier of Familial Mediterranean Fever (AR) Associated gene(s): <i>MEFV</i> Variant(s) Detected: c.2177T>C, p.V726A, Pathogenic, Heterozygous (one copy)</p> <p>Carrier of Phenylalanine Hydroxylase Deficiency (AR) Associated gene(s): <i>PAH</i> Variant(s) Detected: c.898G>T, p.A300S, Pathogenic, Heterozygous (one copy)</p> | <p>Negative for all other genes tested To view a full list of genes and diseases tested please see Table 1 in this report</p> |

AR=Autosomal recessive; XL=X-linked

Recommendations

- Testing the partner for the above positive disorder(s) and genetic counseling are recommended.
- Please note that for female carriers of X-linked diseases, follow-up testing of a male partner is not indicated.
- CGG repeat analysis of *FMR1* for fragile X syndrome is not performed on males as repeat expansion of premutation alleles is not expected in the male germline.
- Individuals of Asian, African, Hispanic and Mediterranean ancestry should also be screened for hemoglobinopathies by CBC and hemoglobin electrophoresis.
- Consideration of residual risk by ethnicity after a negative carrier screen is recommended for the other diseases on the panel, especially in the case of a positive family history for a specific disorder.

Interpretation of positive results

Beta-Globin-Related Hemoglobinopathies (AR)

Results and Interpretation

A heterozygous (one copy) likely pathogenic missense variant, c.208G>A, p.G70S (Hb City of Hope), was detected in the *HBB* gene (NM_000518.4). Please note that this variant has been reported to cause a thalassemia phenotype when found in trans with beta (0) variants, and has also been reported to cause disease when identified in trans with HbS (PMID: 2467892, 25113778 and 21302591). Carriers of this variant do not display any hematological manifestations of beta-thalassemia and have normal CBC and hemoglobin electrophoresis; however, they are considered to be carriers of beta-thalassemia. Beta-thalassemia patients who are homozygous for this variant have not been reported in the literature; therefore, this variant may not cause a disease phenotype when homozygous. When this variant is present in trans with a pathogenic variant, it is considered to be causative for beta-globin-related hemoglobinopathies. Therefore, this individual is expected to be at least a carrier for beta-globin-related hemoglobinopathies.

What is Beta-Globin-Related Hemoglobinopathies?

Pathogenic variants in the beta-globin gene (*HBB*) cause a variety of autosomal recessive diseases of aberrant hemoglobin, the protein that carries oxygen in the blood. The most frequent hemoglobinopathies are beta-thalassemia, sickle cell disease and HbC disease.

- In individuals with beta-thalassemia, hemoglobin is not properly synthesized and results in small red blood cells that are inefficient at carrying oxygen. Individuals with severe beta-thalassemia require life-long blood transfusions and chelation therapy to remove the extra iron that results from the blood transfusions. Individuals with milder forms of beta-thalassemia may not require transfusions. Life expectancy may be shortened due to cardiac complications of iron overload. Individuals carrying one pathogenic allele causing beta-thalassemia in addition to 5 or more copies of HBA may develop a thalassemia intermedia phenotype with a variable clinical presentation, and may require recurrent transfusions.
- Sickle cell disease is caused by the inheritance of two copies of Hemoglobin S (HbS), encoded by a specific *HBB* variant. Symptoms typically first present in infancy or childhood and include chronic anemia, pain and/or swelling in the hands and feet, episodes of severe pain, and infections. The clinical presentation is highly variable between affected individuals. The life expectancy for individuals with sickle cell disease may be shortened. HbS can also cause related diseases if it is inherited along with a different type of variant in *HBB*.
- HbC disease is caused by the inheritance of two copies of Hemoglobin C (HbC), encoded by a specific *HBB* variant. HbC disease causes mild anemia in some patients, but the majority of affected individuals do not have any symptoms and have a normal life expectancy. HbC can also cause disease if it is inherited with another type of abnormal hemoglobin, the most common being HbS. The inheritance of one copy each of HbS and HbC result in SC disease, which may cause chronic anemia, pain and/or swelling in the hands and feet, episodes of severe pain, infections, and retinal disease. The life expectancy for individuals with SC disease may be shortened.

The type of disease that will develop can be predicted based on the variants inherited. Variants causing beta-thalassemia are prevalent in Mediterranean and South-East Asian populations, whereas HbS is most common in people of African, Mediterranean, Middle Eastern, and Indian ancestry. HbC is most common in people of African descent.

Congenital Adrenal Hyperplasia due to 21-Hydroxylase Deficiency (AR)

Results and Interpretation

CYP21A2 copy number: 2

No pathogenic copy number variants detected

CYP21A2 sequencing: c.841G>T, p.V281L, Pathogenic, Heterozygous (one copy)

Genes analyzed: *CYP21A2* (NM_000500.6)

Inheritance: Autosomal Recessive

A heterozygous (one copy) pathogenic missense variant, c.841G>T, p.V281L, was detected in the *CYP21A2* gene (NM_000500.6). Please note that this variant is typically causative for the non-classic form of congenital adrenal hyperplasia (PMID: 29450859). Variants associated with the non-classic form usually cause non-classic congenital adrenal hyperplasia when found in trans with a pathogenic allele, regardless of whether the second variant is associated with classic or non-classic disease (PMID: 29450859). Therefore, this individual is expected to be at least a carrier for non-classic congenital adrenal hyperplasia. Heterozygous carriers are not expected to exhibit symptoms of this disease.

What is congenital adrenal hyperplasia (due to 21-hydroxylase deficiency)?

Congenital adrenal hyperplasia (CAH) is a group of autosomal recessive disorders resulting from deficiency in the enzymes involved in cortisol biosynthesis. The majority (95%) of CAH cases are due to 21-hydroxylase deficiency (21-OHD CAH), which is caused by homozygous or compound heterozygous pathogenic variants in the gene *CYP21A2*. Approximately 20% of mutant alleles have deletions of 30 kb that have been generated by unequal meiotic crossing-over between the two genes. Another 75% of mutant alleles are due to gene conversion events, where an inactivating mutation from the *CYP21A1P* pseudogene is introduced into one copy of the *CYP21A2* gene, thus making the gene non-functional. Three different forms of 21-OHD CAH have been reported: a classic salt wasting form, a classic simple virilizing form, and a non-classic form.

- The classic salt wasting form results from a nonfunctional enzyme and is the most severe. The phenotype includes prenatal onset of virilization and inadequate adrenal aldosterone secretion that can result in fatal salt-wasting crises.
- The classic simple virilizing form results from low levels of functional enzyme and involves prenatal virilization but no salt-wasting.
- The non-classic form, which results from a mild enzyme deficiency, occurs postnatally and involves phenotypes associated with hyperandrogenism, such as hirsutism, delayed menarche, and infertility.

Treatment for the classic forms of the disorder include glucocorticoid and mineralocorticoid replacement therapy, as well as the possibility of feminizing genitoplasty, while patients with the non-classic form usually do not require treatment. The life expectancy for this disorder can be normal with treatment, however the occurrence of salt-wasting crises can be fatal.

Familial Mediterranean Fever (AR)

Results and Interpretation

A heterozygous (one copy) pathogenic missense variant, c.2177T>C, p.V726A, was detected in the *MEFV* gene (NM_000243.2). When this variant is present in trans with a pathogenic variant, it is considered to be causative for familial Mediterranean fever. Therefore, this individual is expected to be at least a carrier for familial Mediterranean fever. Heterozygous carriers are usually asymptomatic, but have occasionally been reported to exhibit mild to severe symptoms of this disease.

What is Familial Mediterranean Fever?

Familial Mediterranean fever is an autosomal recessive disorder caused by pathogenic variants in the gene *MEFV*. It is particularly common in Middle Eastern and Mediterranean populations, as well as individuals of Ashkenazi or Sephardic Jewish ancestry. Clinical symptoms are variable, with some patients having mild forms and never requiring clinical attention. Two main forms of the disease exist:

- Type 1: Recurrent bouts of fever, inflammation and pain in the abdomen or the joints. Depending on the individual, these bouts may occur often or rarely. Each episode typically lasts about 3 days. Some patients have symptoms of discomfort before an episode begins.
- Type 2: Some patients who do not experience fever episodes may develop a buildup of proteins called amyloids in the kidneys. This can lead to kidney damage and end-stage renal disease, requiring dialysis or kidney transplant.

Life expectancy is not reduced, except in untreated patients with severe kidney manifestations. Certain variants are associated with more severe disease, development of amyloidosis, and earlier onset of symptoms.

Phenylalanine Hydroxylase Deficiency (AR)

Results and Interpretation

A heterozygous (one copy) pathogenic missense variant, c.898G>T, p.A300S, was detected in the *PAH* gene (NM_000277.1). When this variant is present in trans with a pathogenic variant, it is considered to be causative for phenylalanine hydroxylase deficiency. Therefore, this individual is expected to be at least a carrier for phenylalanine hydroxylase deficiency. Heterozygous carriers are not expected to exhibit symptoms of this disease.

What is Phenylalanine Hydroxylase Deficiency?

Phenylalanine hydroxylase deficiency is an autosomal recessive disorder caused by pathogenic variants in the gene *PAH*. While it is found in many different ethnicities, it is particularly prevalent in Sephardic Jewish, Sicilian, Irish, and Turkish individuals, as well as Caucasians. Pathogenic *PAH* variants result in loss of function of the phenylalanine hydroxylase enzyme, which breaks down the amino acid phenylalanine. The most severe form of the disease is called phenylketonuria. If untreated, buildup of phenylalanine will result in irreversible brain damage and severe intellectual disability. Treatment involves the removal of phenylalanine from the diet. Even with strict adherence to the treatment, some neurologic deficiencies have been noticed in long-term survivors. Psychological problems, including anxiety, depression, phobias and panic attacks may occur in adults who do not comply well to their treatment. Some patients have a milder form of hyperphenylalaninemia and may

tolerate higher levels of phenylalanine in their diet. Depending on the genotype, patients may be responsive to BH4, which can direct their treatment. However, it is not always possible to predict the severity of the disease based on genotype.

Test description

This patient was tested for a panel of diseases using a combination of sequencing, targeted genotyping and copy number analysis. Please note that negative results reduce but do not eliminate the possibility that this individual is a carrier for one or more of the disorders tested. Please see Table 1 for a list of genes and diseases tested, and go.sema4.com/residualrisk for specific detection rates and residual risk by ethnicity. With individuals of mixed ethnicity, it is recommended to use the highest residual risk estimate. Only variants determined to be pathogenic or likely pathogenic are reported in this carrier screening test.



Christie Buchovecky, Ph.D., Assistant Director, Reproductive Genomic

Laboratory Medical Consultant: George A. Diaz, M.D., Ph.D

Genes and diseases tested

For specific detection rates and residual risk by ethnicity, please visit go.sema4.com/residualrisk

Table 1: List of genes and diseases tested with detailed results

| Disease | Gene | Inheritance Pattern | Status | Detailed Summary |
|---|------------------|---------------------|--------------|---|
| ⊕ Positive | | | | |
| Beta-Globin-Related Hemoglobinopathies | <i>HBB</i> | AR | Carrier | c.208G>A, p.G70S (Hb City of Hope), Likely Pathogenic, Heterozygous (one copy) |
| Congenital Adrenal Hyperplasia due to 21-Hydroxylase Deficiency | <i>CYP21A2</i> | AR | Carrier | <i>CYP21A2</i> copy number: 2 No pathogenic copy number variants detected <i>CYP21A2</i> sequencing: c.841G>T, p.V281L, Pathogenic, Heterozygous (one copy) |
| Familial Mediterranean Fever | <i>MEFV</i> | AR | Carrier | c.2177T>C, p.V726A, Pathogenic, Heterozygous (one copy) |
| Phenylalanine Hydroxylase Deficiency | <i>PAH</i> | AR | Carrier | c.898G>T, p.A300S, Pathogenic, Heterozygous (one copy) |
| ⊖ Negative | | | | |
| 3-Beta-Hydroxysteroid Dehydrogenase Type II Deficiency | <i>HSD3B2</i> | AR | Reduced Risk | |
| 3-Methylcrotonyl-CoA Carboxylase Deficiency (MCCC1-Related) | <i>MCCC1</i> | AR | Reduced Risk | |
| 3-Methylcrotonyl-CoA Carboxylase Deficiency (MCCC2-Related) | <i>MCCC2</i> | AR | Reduced Risk | |
| 3-Methylglutaconic Aciduria, Type III | <i>OPA3</i> | AR | Reduced Risk | |
| 3-Phosphoglycerate Dehydrogenase Deficiency | <i>PHGDH</i> | AR | Reduced Risk | |
| 6-Pyruvoyl-Tetrahydropterin Synthase Deficiency | <i>PTS</i> | AR | Reduced Risk | |
| Abetalipoproteinemia | <i>MTTP</i> | AR | Reduced Risk | |
| Achromatopsia (CNGB3-related) | <i>CNGB3</i> | AR | Reduced Risk | |
| Acrodermatitis Enteropathica | <i>SLC39A4</i> | AR | Reduced Risk | |
| Acute Infantile Liver Failure | <i>TRMU</i> | AR | Reduced Risk | |
| Acyl-CoA Oxidase I Deficiency | <i>ACOX1</i> | AR | Reduced Risk | |
| Adenosine Deaminase Deficiency | <i>ADA</i> | AR | Reduced Risk | |
| Adrenoleukodystrophy, X-Linked | <i>ABCD1</i> | XL | Reduced Risk | |
| Aicardi-Goutieres Syndrome (SAMHD1-Related) | <i>SAMHD1</i> | AR | Reduced Risk | |
| Alpha-Mannosidosis | <i>MAN2B1</i> | AR | Reduced Risk | |
| Alpha-Thalassemia | <i>HBA1/HBA2</i> | AR | Reduced Risk | <i>HBA1</i> Copy Number: 2 <i>HBA2</i> Copy Number: 2 No pathogenic copy number variants detected <i>HBA1/HBA2</i> Sequencing: Negative |
| Alpha-Thalassemia Intellectual Disability Syndrome | <i>ATRX</i> | XL | Reduced Risk | |
| Alport Syndrome (COL4A3-Related) | <i>COL4A3</i> | AR | Reduced Risk | |
| Alport Syndrome (COL4A4-Related) | <i>COL4A4</i> | AR | Reduced Risk | |
| Alport Syndrome (COL4A5-Related) | <i>COL4A5</i> | XL | Reduced Risk | |
| Alstrom Syndrome | <i>ALMS1</i> | AR | Reduced Risk | |
| Andermann Syndrome | <i>SLC12A6</i> | AR | Reduced Risk | |
| Argininosuccinic Aciduria | <i>ASL</i> | AR | Reduced Risk | |
| Aromatase Deficiency | <i>CYP19A1</i> | AR | Reduced Risk | |
| Arthrogryposis, Mental Retardation, and Seizures | <i>SLC35A3</i> | AR | Reduced Risk | |
| Asparagine Synthetase Deficiency | <i>ASNS</i> | AR | Reduced Risk | |
| Aspartylglycosaminuria | <i>AGA</i> | AR | Reduced Risk | |
| Ataxia With Isolated Vitamin E Deficiency | <i>TTPA</i> | AR | Reduced Risk | |
| Ataxia-Telangiectasia | <i>ATM</i> | AR | Reduced Risk | |
| Autosomal Recessive Spastic Ataxia of Charlevoix-Saguenay | <i>SACS</i> | AR | Reduced Risk | |
| Bardet-Biedl Syndrome (BBS10-Related) | <i>BBS10</i> | AR | Reduced Risk | |

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|---|-----------------|----|--------------|
| Bardet-Biedl Syndrome (<i>BBS12</i> -Related) | <i>BBS12</i> | AR | Reduced Risk |
| Bardet-Biedl Syndrome (<i>BBS1</i> -Related) | <i>BBS1</i> | AR | Reduced Risk |
| Bardet-Biedl Syndrome (<i>BBS2</i> -Related) | <i>BBS2</i> | AR | Reduced Risk |
| Bare Lymphocyte Syndrome, Type II | <i>CLTA</i> | AR | Reduced Risk |
| Bartter Syndrome, Type 4A | <i>BSND</i> | AR | Reduced Risk |
| Bernard-Soulier Syndrome, Type A1 | <i>GP1BA</i> | AR | Reduced Risk |
| Bernard-Soulier Syndrome, Type C | <i>GP9</i> | AR | Reduced Risk |
| Beta-Ketothiolase Deficiency | <i>ACAT1</i> | AR | Reduced Risk |
| Bilateral Frontoparietal Polymicrogyria | <i>GPR56</i> | AR | Reduced Risk |
| Biotinidase Deficiency | <i>BTB</i> | AR | Reduced Risk |
| Bloom Syndrome | <i>BLM</i> | AR | Reduced Risk |
| Canavan Disease | <i>ASPA</i> | AR | Reduced Risk |
| Carbamoylphosphate Synthetase I Deficiency | <i>CPS1</i> | AR | Reduced Risk |
| Carnitine Palmitoyltransferase IA Deficiency | <i>CPT1A</i> | AR | Reduced Risk |
| Carnitine Palmitoyltransferase II Deficiency | <i>CPT2</i> | AR | Reduced Risk |
| Carpenter Syndrome | <i>RAB23</i> | AR | Reduced Risk |
| Cartilage-Hair Hypoplasia | <i>RMRP</i> | AR | Reduced Risk |
| Cerebral Creatine Deficiency Syndrome 1 | <i>SLC6A8</i> | XL | Reduced Risk |
| Cerebral Creatine Deficiency Syndrome 2 | <i>GAMT</i> | AR | Reduced Risk |
| Cerebrotendinous Xanthomatosis | <i>CYP27A1</i> | AR | Reduced Risk |
| Charcot-Marie-Tooth Disease, Type 4D | <i>NDRG1</i> | AR | Reduced Risk |
| Charcot-Marie-Tooth Disease, Type 5 / Arts Syndrome | <i>PRPS1</i> | XL | Reduced Risk |
| Charcot-Marie-Tooth Disease, X-Linked | <i>GJB1</i> | XL | Reduced Risk |
| Choreoacanthocytosis | <i>VPS13A</i> | AR | Reduced Risk |
| Choroideremia | <i>CHM</i> | XL | Reduced Risk |
| Chronic Granulomatous Disease (<i>CYBA</i> -Related) | <i>CYBA</i> | AR | Reduced Risk |
| Chronic Granulomatous Disease (<i>CYBB</i> -Related) | <i>CYBB</i> | XL | Reduced Risk |
| Citrin Deficiency | <i>SLC25A13</i> | AR | Reduced Risk |
| Citrullinemia, Type 1 | <i>ASS1</i> | AR | Reduced Risk |
| Cohen Syndrome | <i>VPS13B</i> | AR | Reduced Risk |
| Combined Malonic and Methylmalonic Aciduria | <i>ACSF3</i> | AR | Reduced Risk |
| Combined Oxidative Phosphorylation Deficiency 1 | <i>GFM1</i> | AR | Reduced Risk |
| Combined Oxidative Phosphorylation Deficiency 3 | <i>TSFM</i> | AR | Reduced Risk |
| Combined Pituitary Hormone Deficiency 2 | <i>PROP1</i> | AR | Reduced Risk |
| Combined Pituitary Hormone Deficiency 3 | <i>LHX3</i> | AR | Reduced Risk |
| Combined SAP Deficiency | <i>PSAP</i> | AR | Reduced Risk |
| Congenital Adrenal Hyperplasia due to 17-Alpha-Hydroxylase Deficiency | <i>CYP17A1</i> | AR | Reduced Risk |
| Congenital Amegakaryocytic Thrombocytopenia | <i>MPL</i> | AR | Reduced Risk |
| Congenital Disorder of Glycosylation, Type Ia | <i>PMM2</i> | AR | Reduced Risk |
| Congenital Disorder of Glycosylation, Type Ib | <i>MPI</i> | AR | Reduced Risk |
| Congenital Disorder of Glycosylation, Type Ic | <i>ALG6</i> | AR | Reduced Risk |
| Congenital Insensitivity to Pain with Anhidrosis | <i>NTRK1</i> | AR | Reduced Risk |
| Congenital Myasthenic Syndrome (<i>CHRNE</i> -Related) | <i>CHRNE</i> | AR | Reduced Risk |
| Congenital Myasthenic Syndrome (<i>RAPSN</i> -Related) | <i>RAPSN</i> | AR | Reduced Risk |
| Congenital Neutropenia (<i>HAX1</i> -Related) | <i>HAX1</i> | AR | Reduced Risk |
| Congenital Neutropenia (<i>VPS45</i> -Related) | <i>VPS45</i> | AR | Reduced Risk |
| Corneal Dystrophy and Perceptive Deafness | <i>SLC4A11</i> | AR | Reduced Risk |
| Corticosterone Methyloxidase Deficiency | <i>CYP11B2</i> | AR | Reduced Risk |
| Cystic Fibrosis | <i>CFTR</i> | AR | Reduced Risk |
| Cystinosis | <i>CTNS</i> | AR | Reduced Risk |
| D-Bifunctional Protein Deficiency | <i>HSD17B4</i> | AR | Reduced Risk |
| Deafness, Autosomal Recessive 77 | <i>LOXHD1</i> | AR | Reduced Risk |
| Duchenne Muscular Dystrophy / Becker Muscular Dystrophy | <i>DMD</i> | XL | Reduced Risk |
| Dyskeratosis Congenita (<i>RTEL1</i> -Related) | <i>RTEL1</i> | AR | Reduced Risk |
| Dystrophic Epidermolysis Bullosa | <i>COL7A1</i> | AR | Reduced Risk |
| Ehlers-Danlos Syndrome, Type VIIC | <i>ADAMTS2</i> | AR | Reduced Risk |
| Ellis-van Creveld Syndrome (<i>EVC</i> -Related) | <i>EVC</i> | AR | Reduced Risk |

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|---|----------|----|--------------|--|
| Emery-Dreifuss Myopathy 1 | EMD | XL | Reduced Risk | |
| Enhanced S-Cone Syndrome | NR2E3 | AR | Reduced Risk | |
| Ethylmalonic Encephalopathy | ETHE1 | AR | Reduced Risk | |
| Fabry Disease | GLA | XL | Reduced Risk | |
| Factor IX Deficiency | F9 | XL | Reduced Risk | |
| Factor XI Deficiency | F11 | AR | Reduced Risk | |
| Familial Autosomal Recessive Hypercholesterolemia | LDLRAP1 | AR | Reduced Risk | |
| Familial Dysautonomia | IKBKAP | AR | Reduced Risk | |
| Familial Hypercholesterolemia | LDLR | AR | Reduced Risk | |
| Familial Hyperinsulinism (ABCC8-Related) | ABCC8 | AR | Reduced Risk | |
| Familial Hyperinsulinism (KCNJ11-Related) | KCNJ11 | AR | Reduced Risk | |
| Fanconi Anemia, Group A | FANCA | AR | Reduced Risk | |
| Fanconi Anemia, Group C | FANCC | AR | Reduced Risk | |
| Fanconi Anemia, Group G | FANCG | AR | Reduced Risk | |
| Fragile X Syndrome | FMR1 | XL | Reduced Risk | FMR1 CGG repeat sizes: Not Performed FMR1 Sequencing: Negative Fragile X CGG triplet repeat expansion testing was not performed at this time, as the patient has either been previously tested or is a male. |
| Fumarase Deficiency | FH | AR | Reduced Risk | |
| GRACILE Syndrome and Other BCS1L-Related Disorders | BCS1L | AR | Reduced Risk | |
| Galactokinase Deficiency | GALK1 | AR | Reduced Risk | |
| Galactosemia | GALT | AR | Reduced Risk | |
| Gaucher Disease | GBA | AR | Reduced Risk | |
| Gitelman Syndrome | SLC12A3 | AR | Reduced Risk | |
| Glutaric Acidemia, Type I | GCDH | AR | Reduced Risk | |
| Glutaric Acidemia, Type IIa | ETFA | AR | Reduced Risk | |
| Glutaric Acidemia, Type IIc | ETFDH | AR | Reduced Risk | |
| Glycine Encephalopathy (AMT-Related) | AMT | AR | Reduced Risk | |
| Glycine Encephalopathy (GLDC-Related) | GLDC | AR | Reduced Risk | |
| Glycogen Storage Disease, Type II | GAA | AR | Reduced Risk | |
| Glycogen Storage Disease, Type III | AGL | AR | Reduced Risk | |
| Glycogen Storage Disease, Type IV / Adult Polyglucosan Body Disease | GBE1 | AR | Reduced Risk | |
| Glycogen Storage Disease, Type Ia | G6PC | AR | Reduced Risk | |
| Glycogen Storage Disease, Type Ib | SLC37A4 | AR | Reduced Risk | |
| Glycogen Storage Disease, Type V | PYGM | AR | Reduced Risk | |
| Glycogen Storage Disease, Type VII | PFKM | AR | Reduced Risk | |
| HMG-CoA Lyase Deficiency | HMGCL | AR | Reduced Risk | |
| Hemochromatosis, Type 2A | HFE2 | AR | Reduced Risk | |
| Hemochromatosis, Type 3 | TFR2 | AR | Reduced Risk | |
| Hereditary Fructose Intolerance | ALDOB | AR | Reduced Risk | |
| Hereditary Spastic Paraparesis 49 | TECPR2 | AR | Reduced Risk | |
| Hermansky-Pudlak Syndrome, Type 1 | HPS1 | AR | Reduced Risk | |
| Hermansky-Pudlak Syndrome, Type 3 | HPS3 | AR | Reduced Risk | |
| Holocarboxylase Synthetase Deficiency | HLCS | AR | Reduced Risk | |
| Homocystinuria (CBS-Related) | CBS | AR | Reduced Risk | |
| Homocystinuria due to MTHFR Deficiency | MTHFR | AR | Reduced Risk | |
| Homocystinuria, cblE Type | MTRR | AR | Reduced Risk | |
| Hydroletharus Syndrome | HYLS1 | AR | Reduced Risk | |
| Hyperornithinemia-Hyperammonemia-Homocitrullinuria Syndrome | SLC25A15 | AR | Reduced Risk | |
| Hypohidrotic Ectodermal Dysplasia 1 | EDA | XL | Reduced Risk | |
| Hypophosphatasia | ALPL | AR | Reduced Risk | |
| Inclusion Body Myopathy 2 | GNE | AR | Reduced Risk | |
| Infantile Cerebral and Cerebellar Atrophy | MED17 | AR | Reduced Risk | |
| Isovaleric Acidemia | IVD | AR | Reduced Risk | |
| Joubert Syndrome 2 | TMEM216 | AR | Reduced Risk | |
| Joubert Syndrome 7 / Meckel Syndrome 5 / COACH Syndrome | RPGRIP1L | AR | Reduced Risk | |

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|---|----------------|----|--------------|
| Junctional Epidermolysis Bullosa (<i>LAMA3</i> -Related) | <i>LAMA3</i> | AR | Reduced Risk |
| Junctional Epidermolysis Bullosa (<i>LAMB3</i> -Related) | <i>LAMB3</i> | AR | Reduced Risk |
| Junctional Epidermolysis Bullosa (<i>LAMC2</i> -Related) | <i>LAMC2</i> | AR | Reduced Risk |
| Krabbe Disease | <i>GALC</i> | AR | Reduced Risk |
| Lamellar Ichthyosis, Type 1 | <i>TGM1</i> | AR | Reduced Risk |
| Leber Congenital Amaurosis 10 and Other CEP290-Related Ciliopathies | <i>CEP290</i> | AR | Reduced Risk |
| Leber Congenital Amaurosis 13 | <i>RDH12</i> | AR | Reduced Risk |
| Leber Congenital Amaurosis 2 / Retinitis Pigmentosa 20 | <i>RPE65</i> | AR | Reduced Risk |
| Leber Congenital Amaurosis 5 | <i>LCA5</i> | AR | Reduced Risk |
| Leber Congenital Amaurosis 8 / Retinitis Pigmentosa 12 / Pigmented Paravenous Chorioretinal Atrophy | <i>CRB1</i> | AR | Reduced Risk |
| Leigh Syndrome, French-Canadian Type | <i>LRPPRC</i> | AR | Reduced Risk |
| Lethal Congenital Contracture Syndrome 1 / Lethal Arthrogyposis with Anterior Horn Cell Disease | <i>GLE1</i> | AR | Reduced Risk |
| Leukoencephalopathy with Vanishing White Matter | <i>EIF2B5</i> | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2A | <i>CAPN3</i> | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2B | <i>DYSF</i> | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2C | <i>SGCG</i> | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2D | <i>SGCA</i> | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2E | <i>SGCB</i> | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2I | <i>FKRP</i> | AR | Reduced Risk |
| Lipoamide Dehydrogenase Deficiency | <i>DLA</i> | AR | Reduced Risk |
| Lipoid Adrenal Hyperplasia | <i>STAR</i> | AR | Reduced Risk |
| Lipoprotein Lipase Deficiency | <i>LPL</i> | AR | Reduced Risk |
| Long-Chain 3-Hydroxyacyl-CoA Dehydrogenase Deficiency | <i>HADHA</i> | AR | Reduced Risk |
| Lysinuric Protein Intolerance | <i>SLC7A7</i> | AR | Reduced Risk |
| Maple Syrup Urine Disease, Type 1a | <i>BCKDHA</i> | AR | Reduced Risk |
| Maple Syrup Urine Disease, Type 1b | <i>BCKDHB</i> | AR | Reduced Risk |
| Meckel Syndrome 1 / Bardet-Biedl Syndrome 13 | <i>MKS1</i> | AR | Reduced Risk |
| Medium Chain Acyl-CoA Dehydrogenase Deficiency | <i>ACADM</i> | AR | Reduced Risk |
| Megalencephalic Leukoencephalopathy with Subcortical Cysts | <i>MLC1</i> | AR | Reduced Risk |
| Menkes Disease | <i>ATP7A</i> | XL | Reduced Risk |
| Metachromatic Leukodystrophy | <i>ARSA</i> | AR | Reduced Risk |
| Methylmalonic Acidemia (<i>MMAA</i> -Related) | <i>MMAA</i> | AR | Reduced Risk |
| Methylmalonic Acidemia (<i>MMAB</i> -Related) | <i>MMAB</i> | AR | Reduced Risk |
| Methylmalonic Acidemia (<i>MUT</i> -Related) | <i>MUT</i> | AR | Reduced Risk |
| Methylmalonic Aciduria and Homocystinuria, Cobalamin C Type | <i>MMACHC</i> | AR | Reduced Risk |
| Methylmalonic Aciduria and Homocystinuria, Cobalamin D Type | <i>MMADHC</i> | AR | Reduced Risk |
| Microphthalmia / Anophthalmia | <i>VSX2</i> | AR | Reduced Risk |
| Mitochondrial Complex I Deficiency (<i>ACAD9</i> -Related) | <i>ACAD9</i> | AR | Reduced Risk |
| Mitochondrial Complex I Deficiency (<i>NDUFAF5</i> -Related) | <i>NDUFAF5</i> | AR | Reduced Risk |
| Mitochondrial Complex I Deficiency (<i>NDUFS6</i> -Related) | <i>NDUFS6</i> | AR | Reduced Risk |
| Mitochondrial DNA Depletion Syndrome 6 / Navajo Neurohepatopathy | <i>MPV17</i> | AR | Reduced Risk |
| Mitochondrial Myopathy and Sideroblastic Anemia 1 | <i>PUS1</i> | AR | Reduced Risk |
| Mucopolipidosis II / IIIA | <i>GNPTAB</i> | AR | Reduced Risk |
| Mucopolipidosis III Gamma | <i>GNPTG</i> | AR | Reduced Risk |
| Mucopolipidosis IV | <i>MCOLN1</i> | AR | Reduced Risk |

| | | | |
|---|----------------|----|--------------|
| Mucopolysaccharidosis Type I | <i>IDUA</i> | AR | Reduced Risk |
| Mucopolysaccharidosis Type II | <i>IDS</i> | XL | Reduced Risk |
| Mucopolysaccharidosis Type IIIA | <i>SGSH</i> | AR | Reduced Risk |
| Mucopolysaccharidosis Type IIIB | <i>NAGLU</i> | AR | Reduced Risk |
| Mucopolysaccharidosis Type IIIC | <i>HGSNAT</i> | AR | Reduced Risk |
| Mucopolysaccharidosis Type IIID | <i>GNS</i> | AR | Reduced Risk |
| Mucopolysaccharidosis Type IVb / GM1 Gangliosidosis | <i>GLB1</i> | AR | Reduced Risk |
| Mucopolysaccharidosis type IX | <i>HYAL1</i> | AR | Reduced Risk |
| Mucopolysaccharidosis type VI | <i>ARSB</i> | AR | Reduced Risk |
| Multiple Sulfatase Deficiency | <i>SUMF1</i> | AR | Reduced Risk |
| Muscle-Eye-Brain Disease and Other <i>POMGNT1</i> -Related Congenital Muscular Dystrophy-Dystroglycanopathies | <i>POMGNT1</i> | AR | Reduced Risk |
| Myoneurogastrointestinal Encephalopathy | <i>TYMP</i> | AR | Reduced Risk |
| Myotubular Myopathy 1 | <i>MTM1</i> | XL | Reduced Risk |
| N-Acetylglutamate Synthase Deficiency | <i>NAGS</i> | AR | Reduced Risk |
| Nemaline Myopathy 2 | <i>NEB</i> | AR | Reduced Risk |
| Nephrogenic Diabetes Insipidus, Type II | <i>AQP2</i> | AR | Reduced Risk |
| Nephrotic Syndrome (<i>NPHS1</i> -Related) / Congenital Finnish Nephrosis | <i>NPHS1</i> | AR | Reduced Risk |
| Nephrotic Syndrome (<i>NPHS2</i> -Related) / Steroid-Resistant Nephrotic Syndrome | <i>NPHS2</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>CLN3</i> -Related) | <i>CLN3</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>CLN5</i> -Related) | <i>CLN5</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>CLN6</i> -Related) | <i>CLN6</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>CLN8</i> -Related) | <i>CLN8</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>MFSD8</i> -Related) | <i>MFSD8</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>PPT1</i> -Related) | <i>PPT1</i> | AR | Reduced Risk |
| Neuronal Ceroid-Lipofuscinosis (<i>TPP1</i> -Related) | <i>TPP1</i> | AR | Reduced Risk |
| Niemann-Pick Disease (<i>SMPD1</i> -Related) | <i>SMPD1</i> | AR | Reduced Risk |
| Niemann-Pick Disease, Type C (<i>NPC1</i> -Related) | <i>NPC1</i> | AR | Reduced Risk |
| Niemann-Pick Disease, Type C (<i>NPC2</i> -Related) | <i>NPC2</i> | AR | Reduced Risk |
| Nijmegen Breakage Syndrome | <i>NBN</i> | AR | Reduced Risk |
| Non-Syndromic Hearing Loss (<i>GJB2</i> -Related) | <i>GJB2</i> | AR | Reduced Risk |
| Odonto-Onycho-Dermal Dysplasia / Schopf-Schulz-Passarge Syndrome | <i>WNT10A</i> | AR | Reduced Risk |
| Omenn Syndrome (<i>RAG2</i> -Related) | <i>RAG2</i> | AR | Reduced Risk |
| Omenn Syndrome / Severe Combined Immunodeficiency, Athabaskan-Type | <i>DCLRE1C</i> | AR | Reduced Risk |
| Ornithine Aminotransferase Deficiency | <i>OAT</i> | AR | Reduced Risk |
| Ornithine Transcarbamylase Deficiency | <i>OTC</i> | XL | Reduced Risk |
| Osteopetrosis 1 | <i>TCIRG1</i> | AR | Reduced Risk |
| Pendred Syndrome | <i>SLC26A4</i> | AR | Reduced Risk |
| Polycystic Kidney Disease, Autosomal Recessive | <i>PKHD1</i> | AR | Reduced Risk |
| Polyglandular Autoimmune Syndrome, Type 1 | <i>AIRE</i> | AR | Reduced Risk |
| Pontocerebellar Hypoplasia, Type 1A | <i>VRK1</i> | AR | Reduced Risk |
| Pontocerebellar Hypoplasia, Type 6 | <i>RARS2</i> | AR | Reduced Risk |
| Primary Carnitine Deficiency | <i>SLC22A5</i> | AR | Reduced Risk |
| Primary Ciliary Dyskinesia (<i>DNAH5</i> -Related) | <i>DNAH5</i> | AR | Reduced Risk |
| Primary Ciliary Dyskinesia (<i>DNAI1</i> -Related) | <i>DNAI1</i> | AR | Reduced Risk |
| Primary Ciliary Dyskinesia (<i>DNAI2</i> -Related) | <i>DNAI2</i> | AR | Reduced Risk |
| Primary Hyperoxaluria, Type 1 | <i>AGXT</i> | AR | Reduced Risk |
| Primary Hyperoxaluria, Type 2 | <i>GRHPR</i> | AR | Reduced Risk |
| Primary Hyperoxaluria, Type 3 | <i>HOGA1</i> | AR | Reduced Risk |
| Progressive Cerebello-Cerebral Atrophy | <i>SEPSECS</i> | AR | Reduced Risk |
| Progressive Familial Intrahepatic Cholestasis, Type 2 | <i>ABCB11</i> | AR | Reduced Risk |
| Propionic Acidemia (<i>PCCA</i> -Related) | <i>PCCA</i> | AR | Reduced Risk |
| Propionic Acidemia (<i>PCCB</i> -Related) | <i>PCCB</i> | AR | Reduced Risk |
| Pycnodysostosis | <i>CTSK</i> | AR | Reduced Risk |
| Pyruvate Dehydrogenase E1-Alpha Deficiency | <i>PDHA1</i> | XL | Reduced Risk |
| Pyruvate Dehydrogenase E1-Beta Deficiency | <i>PDHB</i> | AR | Reduced Risk |

| | | | | |
|--|-----------------|----|--------------|---|
| Renal Tubular Acidosis and Deafness | <i>ATP6V1B1</i> | AR | Reduced Risk | |
| Retinitis Pigmentosa 25 | <i>EYS</i> | AR | Reduced Risk | |
| Retinitis Pigmentosa 26 | <i>CERKL</i> | AR | Reduced Risk | |
| Retinitis Pigmentosa 28 | <i>FAM161A</i> | AR | Reduced Risk | |
| Retinitis Pigmentosa 59 | <i>DHDDS</i> | AR | Reduced Risk | |
| Rhizomelic Chondrodysplasia Punctata, Type 1 | <i>PEX7</i> | AR | Reduced Risk | |
| Rhizomelic Chondrodysplasia Punctata, Type 3 | <i>AGPS</i> | AR | Reduced Risk | |
| Roberts Syndrome | <i>ESCO2</i> | AR | Reduced Risk | |
| Salla Disease | <i>SLC17A5</i> | AR | Reduced Risk | |
| Sandhoff Disease | <i>HEXB</i> | AR | Reduced Risk | |
| Schimke Immunoosseous Dysplasia | <i>SMARCAL1</i> | AR | Reduced Risk | |
| Segawa Syndrome | <i>TH</i> | AR | Reduced Risk | |
| Sjogren-Larsson Syndrome | <i>ALDH3A2</i> | AR | Reduced Risk | |
| Smith-Lemli-Opitz Syndrome | <i>DHCR7</i> | AR | Reduced Risk | |
| Spinal Muscular Atrophy | <i>SMN1</i> | AR | Reduced Risk | <i>SMN1</i> copy number: >=3 <i>SMN2</i> copy number: 1 c.*3+80T>G: Detected |
| Spondylothoracic Dysostosis | <i>MESP2</i> | AR | Reduced Risk | |
| Steel Syndrome | <i>COL27A1</i> | AR | Reduced Risk | |
| Stuve-Wiedemann Syndrome | <i>LIFR</i> | AR | Reduced Risk | |
| Sulfate Transporter-Related Osteochondrodysplasia | <i>SLC26A2</i> | AR | Reduced Risk | |
| Tay-Sachs Disease | <i>HEXA</i> | AR | Reduced Risk | Tay-Sachs disease enzyme: Non-carrier White blood cells: Non-carrier <ul style="list-style-type: none"> Hex A%: 62.2% (Non-carrier : 55.0 - 72.0%; Carrier: <50%) Total hexosaminidase activity: 1943 nmoL/hr/mg Plasma: Non-carrier <ul style="list-style-type: none"> Hex A%: 73.4 (Non-carrier : 58.0 - 72.0%; Carrier: <54%) Total hexosaminidase activity: 398 nmoL/hr/ml HEXA Sequencing: Negative |
| Tyrosinemia, Type I | <i>FAH</i> | AR | Reduced Risk | |
| Usher Syndrome, Type IB | <i>MYO7A</i> | AR | Reduced Risk | |
| Usher Syndrome, Type IC | <i>USH1C</i> | AR | Reduced Risk | |
| Usher Syndrome, Type ID | <i>CDH23</i> | AR | Reduced Risk | |
| Usher Syndrome, Type IF | <i>PCDH15</i> | AR | Reduced Risk | |
| Usher Syndrome, Type IIA | <i>USH2A</i> | AR | Reduced Risk | |
| Usher Syndrome, Type III | <i>CLRN1</i> | AR | Reduced Risk | |
| Very Long Chain Acyl-CoA Dehydrogenase Deficiency | <i>ACADVL</i> | AR | Reduced Risk | |
| Walker-Warburg Syndrome and Other <i>FKTN</i> -Related Dystrophies | <i>FKTN</i> | AR | Reduced Risk | |
| Wilson Disease | <i>ATP7B</i> | AR | Reduced Risk | |
| Wolman Disease / Cholesteryl Ester Storage Disease | <i>LIPA</i> | AR | Reduced Risk | |
| X-Linked Juvenile Retinoschisis | <i>RS1</i> | XL | Reduced Risk | |
| X-Linked Severe Combined Immunodeficiency | <i>IL2RG</i> | XL | Reduced Risk | |
| Zellweger Syndrome Spectrum (<i>PEX10</i> -Related) | <i>PEX10</i> | AR | Reduced Risk | |
| Zellweger Syndrome Spectrum (<i>PEX1</i> -Related) | <i>PEX1</i> | AR | Reduced Risk | |
| Zellweger Syndrome Spectrum (<i>PEX2</i> -Related) | <i>PEX2</i> | AR | Reduced Risk | |
| Zellweger Syndrome Spectrum (<i>PEX6</i> -Related) | <i>PEX6</i> | AR | Reduced Risk | |

AR=Autosomal recessive; XL=X-linked

Test methods and comments

Genomic DNA isolated from this patient was analyzed by one or more of the following methodologies, as applicable:

Fragile X CGG Repeat Analysis (Analytical Detection Rate >99%)

PCR amplification using Asuragen, Inc. AmplideX® *FMR1* PCR reagents followed by capillary electrophoresis for allele sizing was performed. Samples positive for *FMR1* CGG repeats in the premutation and full mutation size range were further analyzed by Southern blot analysis to assess the size and methylation status of the *FMR1* CGG repeat.

Genotyping (Analytical Detection Rate >99%)

Multiplex PCR amplification and allele specific primer extension analyses using the MassARRAY® System were used to identify variants that are complex in nature or are present in low copy repeats. Rare sequence variants may interfere with assay performance.

Multiplex Ligation-Dependent Probe Amplification (MLPA) (Analytical Detection Rate >99%)

MLPA® probe sets and reagents from MRC-Holland were used for copy number analysis of specific targets versus known control samples. False positive or negative results may occur due to rare sequence variants in target regions detected by MLPA probes. Analytical sensitivity and specificity of the MLPA method are both 99%.

For alpha thalassemia, the copy numbers of the *HBA1* and *HBA2* genes were analyzed. Alpha-globin gene deletions, triplications, and the Constant Spring (CS) mutation are assessed. This test is expected to detect approximately 90% of all alpha-thalassemia mutations, varying by ethnicity, carriers of alpha-thalassemia with three or more *HBA* copies on one chromosome, and one or no copies on the other chromosome, may not be detected. With the exception of triplications, other benign alpha-globin gene polymorphisms will not be reported. Analyses of *HBA1* and *HBA2* are performed in association with long-range PCR of the coding regions followed by short-read sequencing.

For Duchenne muscular dystrophy, the copy numbers of all *DMD* exons were analyzed. Potentially pathogenic single exon deletions and duplications are confirmed by a second method. Analysis of *DMD* is performed in association with sequencing of the coding regions.

For congenital adrenal hyperplasia, the copy number of the *CYP21A2* gene was analyzed. This analysis can detect large deletions due to unequal meiotic crossing-over between *CYP21A2* and the pseudogene *CYP21A1P*. These 30-kb deletions make up approximately 20% of *CYP21A2* pathogenic alleles. This test may also identify certain point mutations in *CYP21A2* caused by gene conversion events between *CYP21A2* and *CYP21A1P*. Some carriers may not be identified by dosage sensitive methods as this testing cannot detect individuals with two copies (duplication) of the *CYP21A2* gene on one chromosome and loss of *CYP21A2* (deletion) on the other chromosome. Analysis of *CYP21A2* is performed in association with long-range PCR of the coding regions followed by short-read sequencing.

For spinal muscular atrophy (SMA), the copy numbers of the *SMN1* and *SMN2* genes were analyzed. The individual dosage of exons 7 and 8 as well as the combined dosage of exons 1, 4, 6 and 8 of *SMN1* and *SMN2* were assessed. Copy number gains and losses can be detected with this assay. Depending on ethnicity, 6 - 29 % of carriers will not be identified by dosage sensitive methods as this testing cannot detect individuals with two copies (duplication) of the *SMN1* gene on one chromosome and loss of *SMN1* (deletion) on the other chromosome (silent 20 carrier) or individuals that carry an intragenic mutation in *SMN1*. Please also note that 2% of individuals with SMA have an *SMN1* mutation that occurred *de novo*. Typically in these cases, only one parent is an SMA carrier.

The presence of the c.*380T>G (chr5:70,247,901T>G) variant allele in an individual with Ashkenazi Jewish or Asian ancestry is typically indicative of a duplication of *SMN1*. When present in an Ashkenazi Jewish or Asian individual with two copies of *SMN1*, c.*380T>G is likely indicative of a silent (20) carrier. In individuals with two copies of *SMN1* with African American, Hispanic or Caucasian ancestry, the presence or absence of c.*380T>G significantly increases or decreases, respectively, the likelihood of being a silent 20 carrier.

Pathogenic or likely pathogenic sequence variants in exon 7 may be detected during testing for the c.*380T>G variant allele; these will be reported if confirmed to be located in *SMN1* using locus-specific Sanger primers.

MLPA for Gaucher disease (*GBA*), cystic fibrosis (*CFTR*), and non-syndromic hearing loss (*GJB2/GJB6*) will only be performed if indicated for confirmation of detected CNVs. If *GBA* analysis was performed, the copy numbers of exons 1, 3, 4, and 6 - 10 of the *GBA* gene (of 11 exons total) were analyzed. If *CFTR* analysis was performed, the copy numbers of all 27 *CFTR* exons were analyzed. If *GJB2/GJB6* analysis was performed, the copy number of the two *GJB2* exons were analyzed, as well as the presence or absence of the two upstream deletions of the *GJB2* regulatory region, del(*GJB6*-D13S1830) and del(*GJB6*-D13S1854).

Next Generation Sequencing (NGS) (Analytical Detection Rate >95%)

NGS was performed on a panel of genes for the purpose of identifying pathogenic or likely pathogenic variants.

Agilent SureSelect™ QXT technology was used with a custom capture library to target the exonic regions and intron/exon splice junctions of the relevant genes, as well as a number of UTR, intronic or promoter regions that contain previously reported mutations. Samples were pooled and sequenced on the Illumina HiSeq 2500 platform in the Rapid Run mode or the Illumina NovaSeq platform in the Xp workflow, using 100 bp paired-end reads. The sequencing data was analyzed using a custom bioinformatics algorithm designed and validated in house.

The coding exons and splice junctions of the known protein-coding RefSeq genes were assessed for the average depth of coverage (minimum of 20X) and data quality threshold values. Most exons not meeting a minimum of >20X read depth across the exon are further analyzed by Sanger sequencing. Please note that several genomic regions present difficulties in mapping or obtaining read depth >20X. The exons contained

within these regions are noted within Table 1 (as "Exceptions") and will not be reflexed to Sanger sequencing if the mapping quality or coverage is poor. Any variants identified during testing in these regions are confirmed by a second method and reported if determined to be pathogenic or likely pathogenic. However, as there is a possibility of false negative results within these regions, detection rates and residual risks for these genes have been calculated with the presumption that variants in these exons will not be detected, unless included in the MassARRAY® genotyping platform.

This test will detect variants within the exons and the intron-exon boundaries of the target regions. Variants outside these regions may not be detected, including, but not limited to, UTRs, promoters, and deep intronic areas, or regions that fall into the Exceptions mentioned above. This technology may not detect all small insertion/deletions and is not diagnostic for repeat expansions and structural genomic variation. In addition, a mutation(s) in a gene not included on the panel could be present in this patient.

Variant interpretation and classification was performed based on the American College of Medical Genetics Standards and Guidelines for the Interpretation of Sequence Variants (Richards et al, 2015). All potentially pathogenic variants may be confirmed by either a specific genotyping assay or Sanger sequencing, if indicated. Any benign variants, likely benign variants or variants of uncertain significance identified during this analysis will not be reported.

Copy Number Variant Analysis (Analytical Detection Rate >95%)

Large duplications and deletions were called from the relative read depths on an exon-by-exon basis using a custom exome hidden Markov model (XHMM) algorithm. Deletions or duplications determined to be pathogenic or likely pathogenic were confirmed by either a custom array CGH platform, quantitative PCR, or MLPA (depending on CNV size and gene content). While this algorithm is designed to pick up deletions and duplications of 2 or more exons in length, potentially pathogenic single-exon CNVs will be confirmed and reported, if detected.

Exon Array (Confirmation method) (Accuracy >99%)

The customized oligonucleotide microarray (Oxford Gene Technology) is a highly-targeted exon-focused array capable of detecting medically relevant microdeletions and microduplications at a much higher resolution than traditional aCGH methods. Each array matrix has approximately 180,000 60-mer oligonucleotide probes that cover the entire genome. This platform is designed based on human genome NCBI Build 37 (hg19) and the CGH probes are enriched to target the exonic regions of the genes in this panel.

Quantitative PCR (Confirmation method) (Accuracy >99%)

The relative quantification PCR is utilized on a Roche Universal Library Probe (UPL) system, which relates the PCR signal of the target region in one group to another. To test for genomic imbalances, both sample DNA and reference DNA is amplified with primer/probesets that specific to the target region and a control region with known genomic copy number. Relative genomic copy numbers are calculated based on the standard $\Delta\Delta C_t$ formula.

Long-Range PCR (Analytical Detection Rate >99%)

Long-range PCR was performed to generate locus-specific amplicons for *CYP21A2*, *HBA1* and *HBA2* and *GBA*. The PCR products were then prepared for short-read NGS sequencing and sequenced. Sequenced reads were mapped back to the original genomic locus and run through the bioinformatics pipeline. If indicated, copy number from MLPA was correlated with the sequencing output to analyze the results. For *CYP21A2*, a certain percentage of healthy individuals carry a duplication of the *CYP21A2* gene, which has no clinical consequences. In cases where two copies of a gene are located on the same chromosome in tandem, only the second copy will be amplified and assessed for potentially pathogenic variants, due to size limitations of the PCR reaction. However, because these alleles contain at least two copies of the *CYP21A2* gene in tandem, it is expected that this patient has at least one functional gene in the tandem allele and this patient is therefore less likely to be a carrier. When an individual carries both a duplication allele and a pathogenic variant, or multiple pathogenic variants, the current analysis may not be able to determine the phase (cis/trans configuration) of the *CYP21A2* alleles identified. Family studies may be required in certain scenarios where phasing is required to determine the carrier status.

Residual Risk Calculations

Carrier frequencies and detection rates for each ethnicity were calculated through the combination of internal curations of >28,000 variants and genomic frequency data from >138,000 individuals across seven ethnic groups in the gnomAD database. Additional variants in HGMD and novel deleterious variants were also incorporated into the calculation. Residual risk values are calculated using a Bayesian analysis combining the *a priori* risk of being a pathogenic mutation carrier (carrier frequency) and the detection rate. They are provided only as a guide for assessing approximate risk given a negative result, and values will vary based on the exact ethnic background of an individual. This report does not represent medical advice but should be interpreted by a genetic counselor, medical geneticist or physician skilled in genetic result interpretation and the relevant medical literature.

Sanger Sequencing (Confirmation method) (Accuracy >99%)

Sanger sequencing, as indicated, was performed using BigDye Terminator chemistry with the ABI 3730 DNA analyzer with target specific amplicons. It also may be used to supplement specific guaranteed target regions that fail NGS sequencing due to poor quality or low depth of

coverage (<20 reads) or as a confirmatory method for NGS positive results. Falsenegative results may occur if rare variants interfere with amplification or annealing.

Tay-Sachs Disease (TSD) Enzyme Analysis (Analytical Detection Rate \geq 98%)

Hexosaminidase activity and Hex A% activity were measured by a standard heat-inactivation, fluorometric method using artificial 4-MU- β -N-acetyl glucosaminide (4-MUG) substrate. This assay is highly sensitive and accurate in detecting Tay-Sachs carriers and individuals affected with TSD. Normal ranges of Hex A% activity are 55.0-72.0 for white blood cells and 58.0-72.0 for plasma. It is estimated that less than 0.5% of Tay-Sachs carriers have non-carrier levels of percent Hex A activity, and therefore may not be identified by this assay. In addition, this assay may detect individuals that are carriers of or are affected with Sandhoff disease. False positive results may occur if benign variants, such as pseudodeficiency alleles, interfere with the enzymatic assay. Falsenegative results may occur if both *HEXA* and *HEXB* pathogenic or pseudodeficiency variants are present in the same individual.

Please note these tests were developed and their performance characteristics were determined by Mount Sinai Genomics, Inc. They have not been cleared or approved by the FDA. These analyses generally provide highly accurate information regarding the patient's carrier or affected status. Despite this high level of accuracy, it should be kept in mind that there are many potential sources of diagnostic error, including misidentification of samples, polymorphisms, or other rare genetic variants that interfere with analysis. Families should understand that rare diagnostic errors may occur for these reasons.

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Variant Classification:

Richards S et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 2015 May;17(5):405-24

Additional disease-specific references available upon request.



| Patient Information | Specimen Information | Client Information |
|---|---|---|
| 556, CB DOB: [REDACTED] AGE: [REDACTED] Gender: M Phone: NG Patient ID: [REDACTED] | Specimen: [REDACTED] Requisition: [REDACTED] Lab Ref #: [REDACTED] Collected: [REDACTED] Received: [REDACTED] Reported: [REDACTED] | Client #: [REDACTED] [REDACTED] GENOMICS, SEMA4 SEMA4 62 SOUTHFIELD AVE STAMFORD, CT 06902-7229 |

Ward: CRYBIO

Cytogenetic Report

CHROMOSOME ANALYSIS, BLOOD - 14596 **Lab:EZ**

CHROMOSOME ANALYSIS, BLOOD

Order ID: [REDACTED]
 Specimen Type: Blood
 Clinical Indication: RULE OUT CHROMOSOME ABNORMALITY

RESULT:
 NORMAL MALE KARYOTYPE

INTERPRETATION:
 Chromosome analysis revealed normal G-band patterns within the limits of standard cytogenetic analysis.

Please expect the results of any other concurrent study in a separate report.

NOMENCLATURE:
 46,XY

ASSAY INFORMATION:

Method: G-Band (Digital Analysis: MetaSyst)
 Cells Counted: 20
 Band Level: 450
 Cells Analyzed: 5
 Cells Karyotyped: 3

This test does not address genetic disorders that cannot be detected by standard cytogenetic methods or rare events such as low level mosaicism or subtle rearrangements.

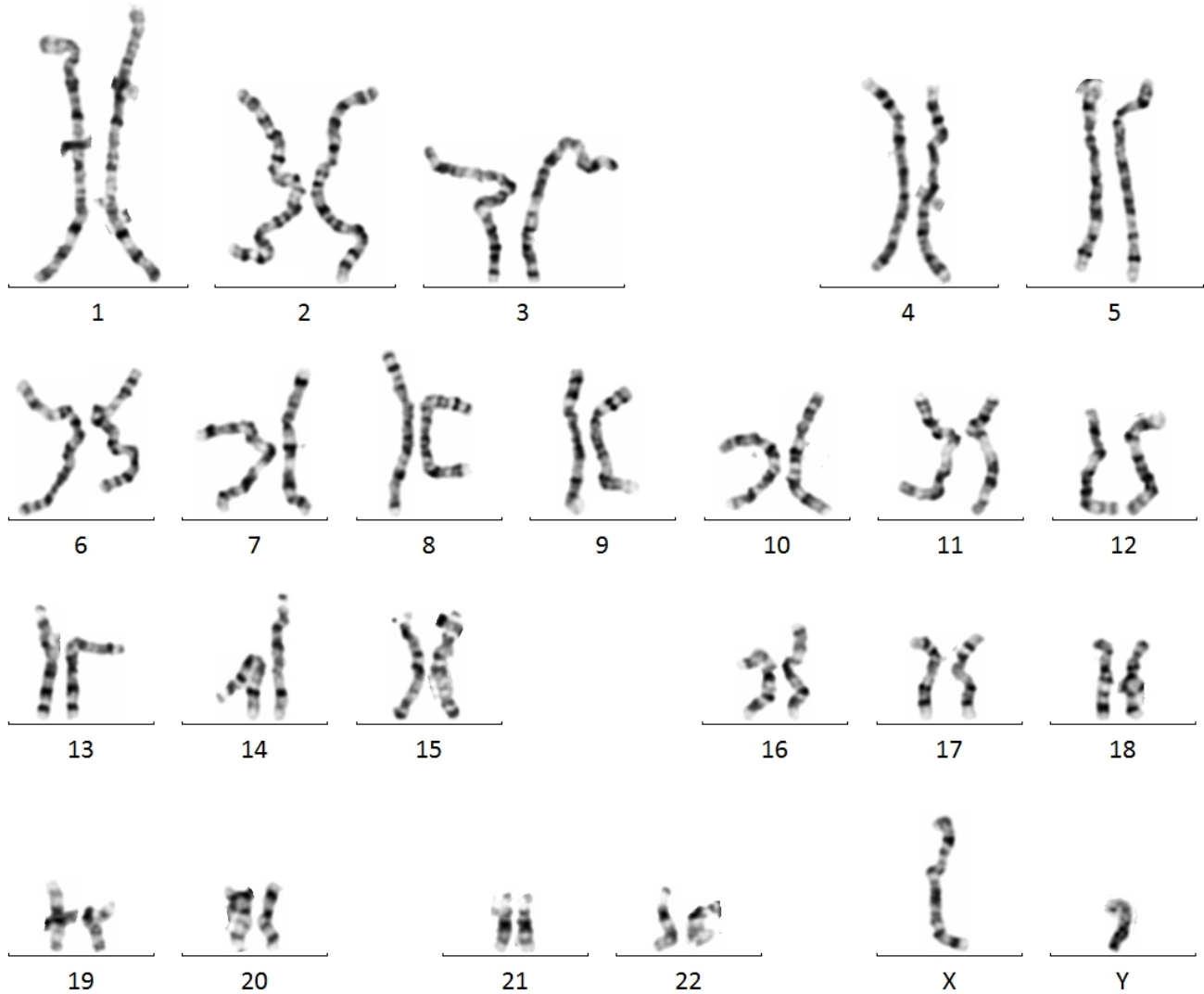
Lakshmi J. Nemana, Ph.D., FACMG

Electronic Signature: [REDACTED]





| Patient Information | Specimen Information | Client Information |
|--|---|---|
| 556, CB DOB: [REDACTED] AGE: [REDACTED] Gender: M Patient ID: [REDACTED] | Specimen: [REDACTED] Collected: [REDACTED] Received: [REDACTED] Reported: [REDACTED] | Client #: [REDACTED] GENOMICS, SEMA4 |



PERFORMING SITE:

EZ QUEST DIAGNOSTICS/NICHOLS SJ, 33608 ORTEGA HWY, SAN JUAN CAPISTRANO, CA 92675-2042 Laboratory Director: IRINA MARAMICA,MD,PHD,MBA, CLIA: 05D0643352

